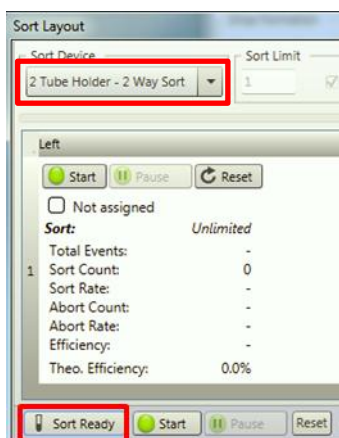


Imaging and Flow Cytometry Core

BD Influx Standard Operation Protocol - Basic Operation

Optimizing the drop phase

1. Verify the 2-tube holder with dummy 15ml tube is in place on the sort stage
2. Select '**2 tube holder – 2 way sort**' as the sort device in the *Sort Layout* pane.
3. Click '**Sort Ready**' in the *Sort Layout* pane to place the dummy in place.



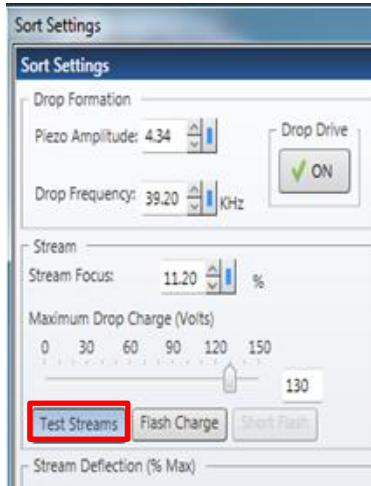
4. Close the deflection plates and press '**Plate**' to charge them

Caution: Risk of electric shock if touch the deflection plates when they are charged.

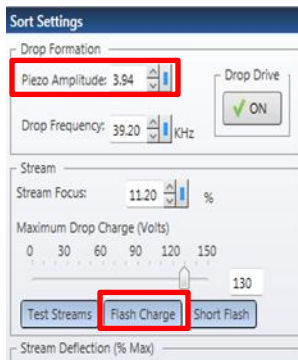


Imaging and Flow Cytometry Core

- Confirm the sort chamber door is closed. Click '**Test Streams**' in the *Sort Setting* pane to initiate test sort.



- Click '**Flash Charge**' and adjust the **Piezo Amplitude** so that the side streams are maximally deflected with no fanning.

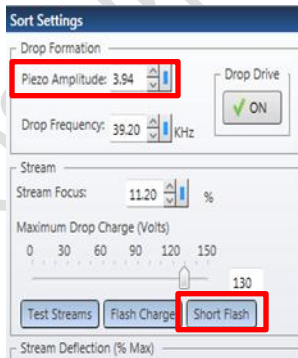


Fanning



No fanning

- Click '**Short Flash**' and fine tune the **Piezo Amplitude** so that the side streams are maximally deflected with no fanning.

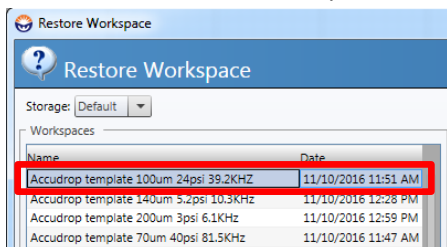


Imaging and Flow Cytometry Core

- Click 'Test Streams' again to turn off.

Accudrop Test

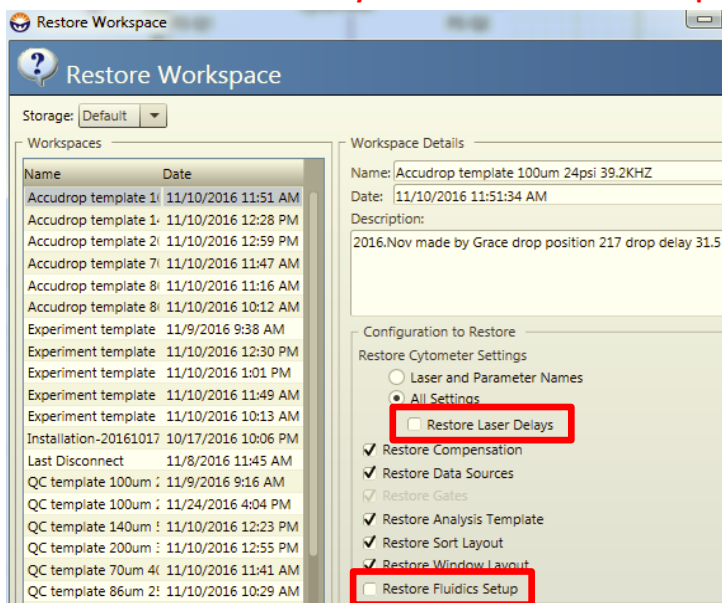
- Click **File > Restore > Workspace**. The following window will appear.



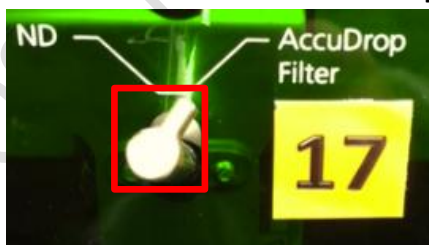
Storage: Choose **Default**

Choose the Accudrop template according to the nozzle size.

- Uncheck "**Restore Laser Delay**" and "**Restore Fluidics Setup**"

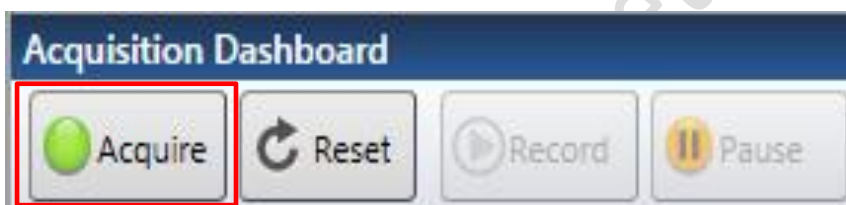


- Turn the **ND** filter knob to **AccuDrop** filter.



Imaging and Flow Cytometry Core

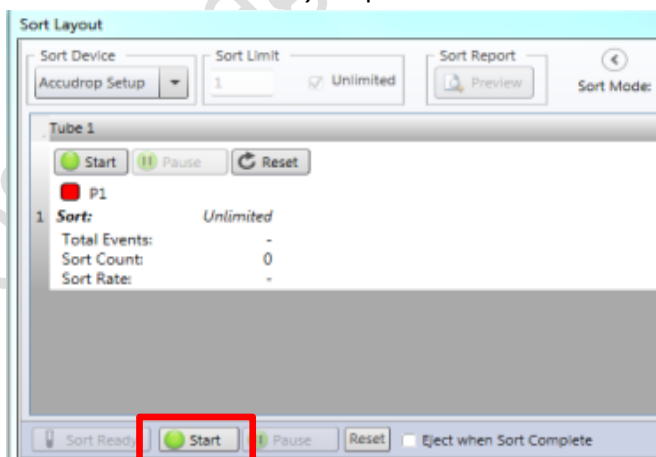
4. Load a tube of AccuDrop beads, press '**Sample**' button beside the sample loading port and click '**Acquire**' on the *Acquisition Dashboard*.



5. Adjust the **Sample Offset** to reach an event rate of between 1000 to 3000 events per second.
(For 100um, event rate should be around 2000 events per second)

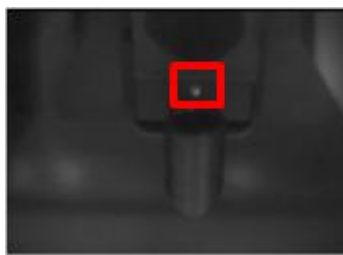
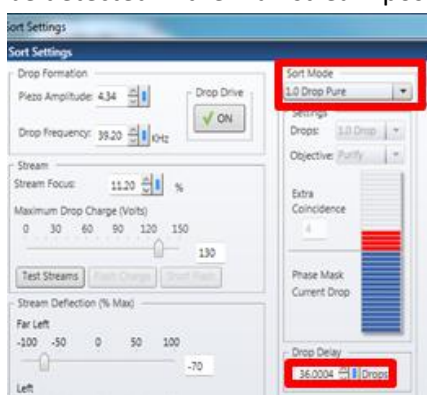


6. Check if '**Plate**' button is on. Turn it on if needed.
7. Click '**Start**' in the *Sort Layout* pane.

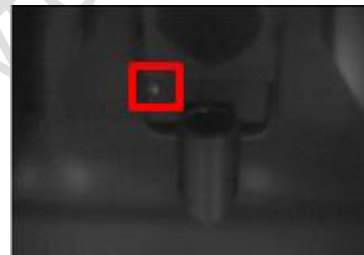


Imaging and Flow Cytometry Core

8. Choose **Sort Mode 2.0 drop Enrich** in the *Sort Settings* pane.
9. Adjust the '**Drop Delay**' until the left stream is the brightest while nearly no fluorescent signals can be detected in the mainstream position.



Wrong Drop Delay



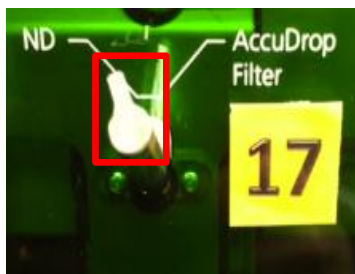
Optimal Drop Delay

10. Choose **Sort Mode 1.0 drop Enrich** in the *Sort Settings* pane and repeat Step 7 to fine tune.
11. Click '**Stop**' in the *Sort Layout* pane and '**Stop**' on the *Acquisition Dashboard*.
12. Press '**Plate**' to turn off the deflection plates.
13. Unload the tube and press '**Backflush**'. Allow the flush for at least 5 seconds.



14. Turn the **Accdrop filter** knob back to **ND** filter position.

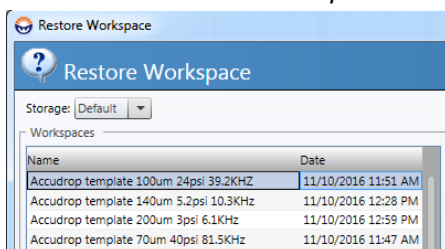
Imaging and Flow Cytometry Core



Attention: If you would like to further culture your collected cells, it is recommended to clean the sample line with FACSClean for 8 minutes at 27.5psi followed by MilliQ Water of same conditions after the Accudrop Test. BSC hood is recommended to be turned on.

Experimental Set up

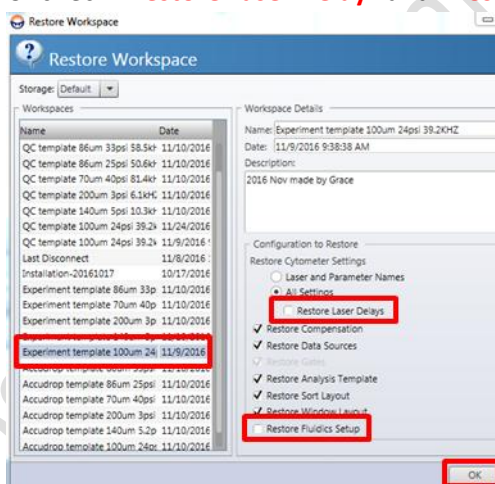
1. Click **File > Restore > Workspace**. The following window will appear.



Storage: Choose **Default**

Choose the Experiment template according to the nozzle size.

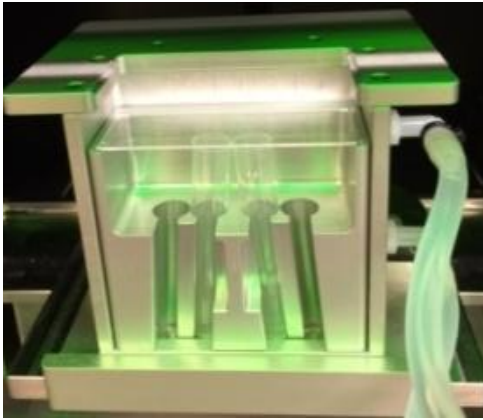
2. Uncheck **"Restore Laser Delay"** and **"Restore Fluidics Setup"**. Click **'OK'**.



3. Place the desired collection device and dummy tubes into the sort chamber and connect the tubes to the ports if cold condition is needed. Place a slide on top of the dummy tubes.



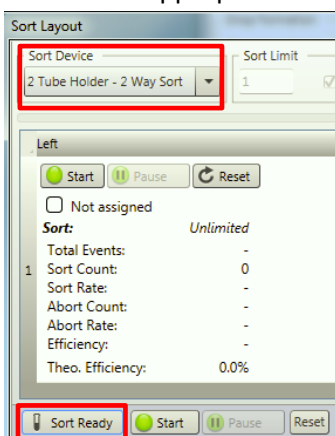
Imaging and Flow Cytometry Core



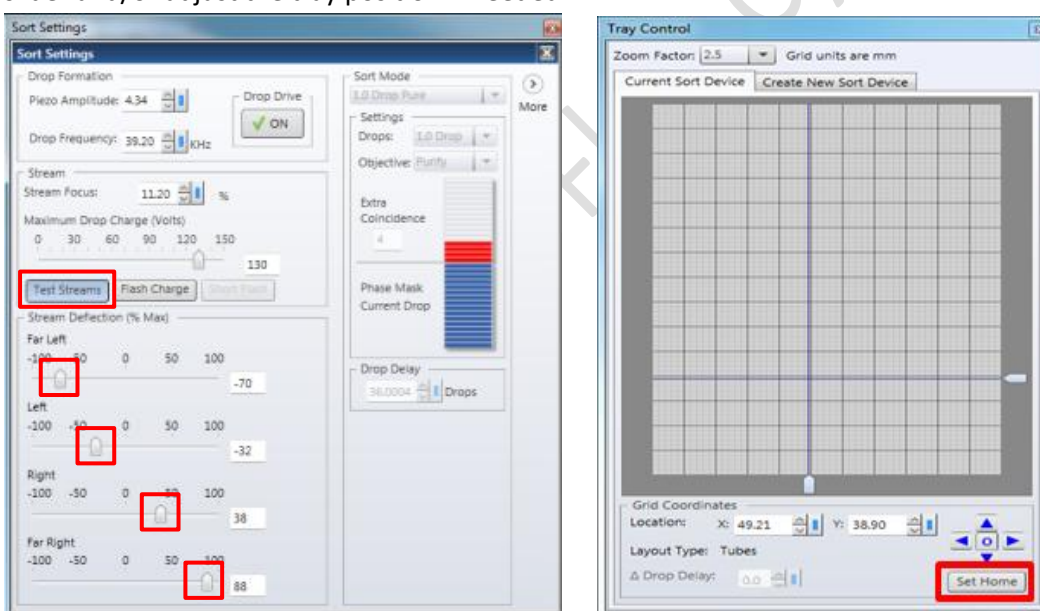
CPoS - Imaging and Flow Cytometry Core

Imaging and Flow Cytometry Core

- Choose the appropriate **Sort Device**. Click '**Sort Ready**' to move the collection device in place.



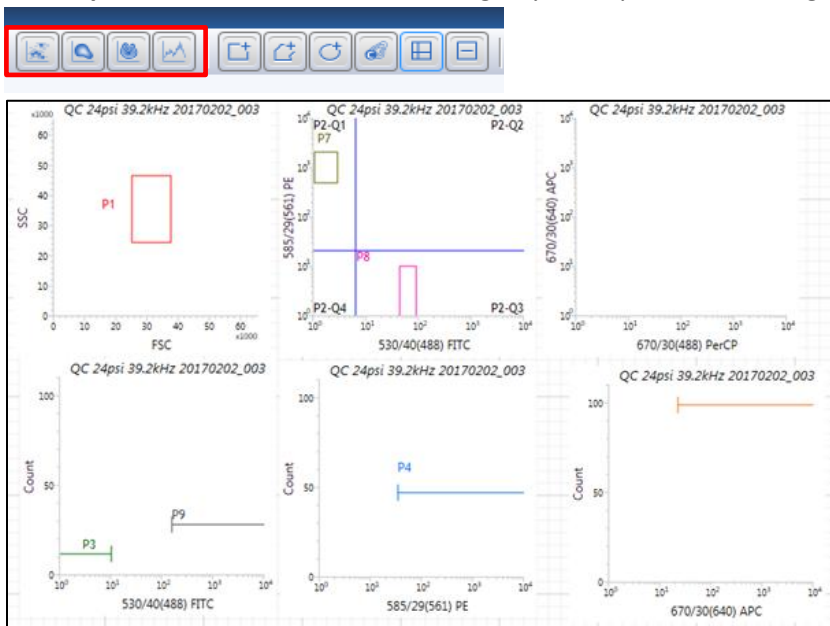
- Press '**Plate**' to charge deflection plates. Double click "**Test Streams**" to place a drop of sheath fluid on the slide to see if the position is correct. Adjust the deflection angle of the side streams using the slider and/or adjust the tray position if needed.



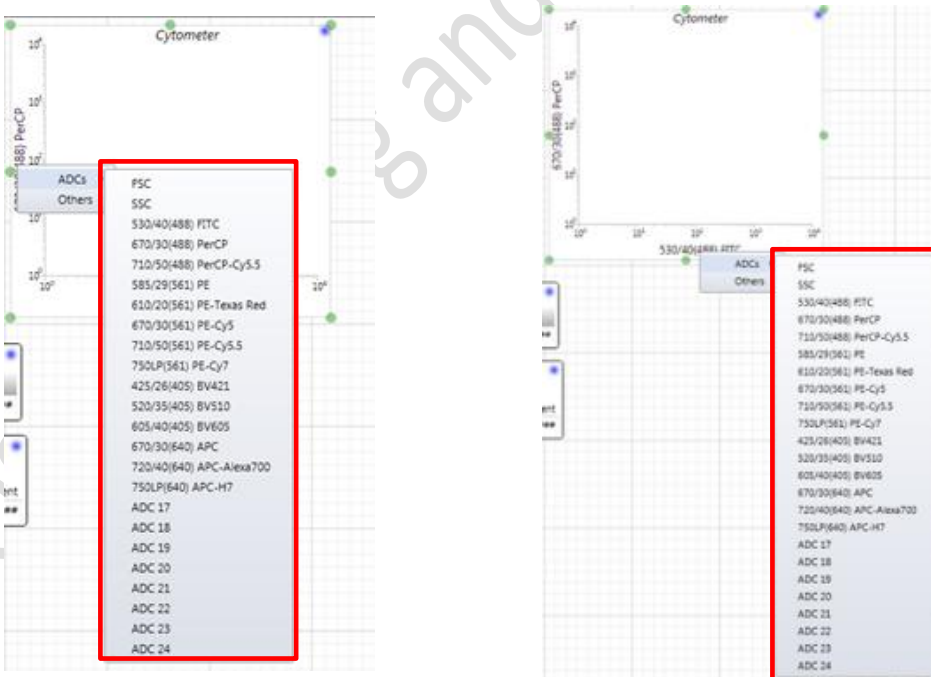
- Remove the slide and Click '**Test Streams**' to verify if the side streams can enter the dummy tubes. Fine tune if needed.
- Click '**Set Home**' in the *Tray Control* pane if the tray position has been modified.
- Press '**Plate**' to turn off the deflection plates.

Imaging and Flow Cytometry Core

- Replace the dummy tubes with the collection tubes that contain enough amount of medium or collection buffer.
- Create **plots** on the *Worksheet* according to your experimental design.

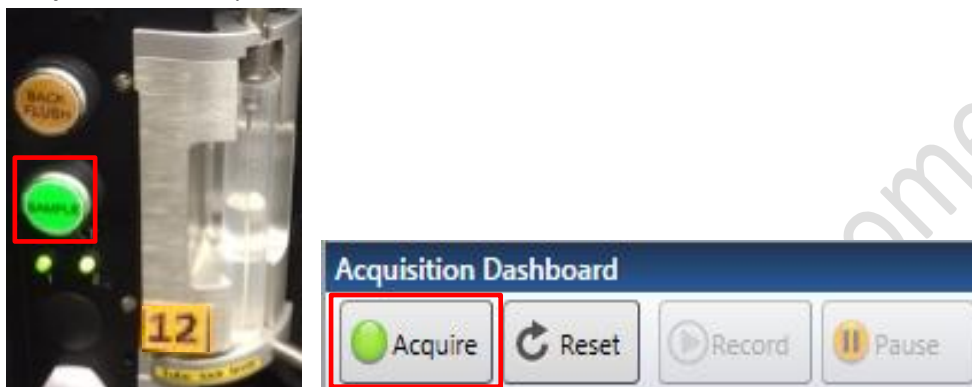


- Right click at X and Y axes label and use ADCs to choose your parameters accordingly.

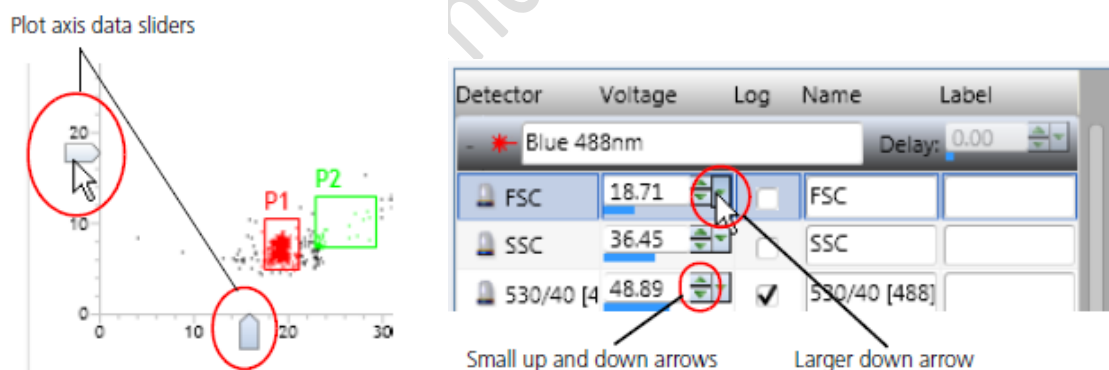


Imaging and Flow Cytometry Core

12. Load a tube of your sample, press '**Sample**' button beside the sample loading port and click '**Acquire**' on the *Acquisition Dashboard*.



13. Adjust the **Sample Offset** and set the sample pressure to around 25psi.
14. Adjust the corresponding PMT voltage using the **slider** on the corresponding axis in a plot or in the *Cytometer Settings* pane.



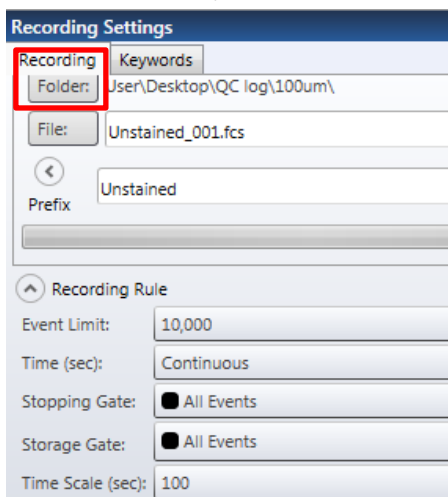
15. Create **Gates** according to your plot type and experiment design. Make sure the Hierarchy of the gates is correct.





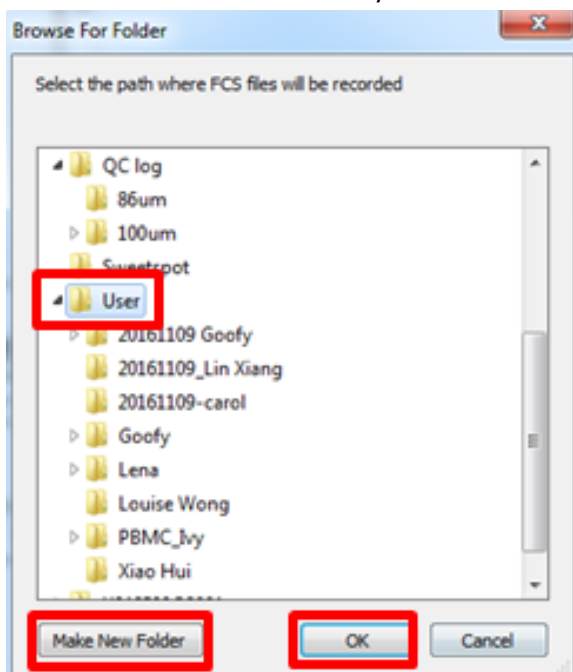
Imaging and Flow Cytometry Core

16. To record FCS file, click the **'Folder'** button in the *Recording Settings* pane.

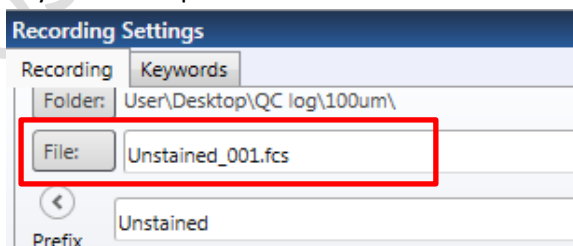


The 'Recording Settings' dialog box is shown. The 'Recording' tab is selected. The 'Folder' button is highlighted with a red box. The 'File' field contains 'Unstained_001.fcs'. The 'Prefix' field contains 'Unstained'. The 'Recording Rule' section is expanded, showing 'Event Limit: 10,000', 'Time (sec): Continuous', 'Stopping Gate: All Events', 'Storage Gate: All Events', and 'Time Scale (sec): 100'.

17. Select **'User'** folder and create yourself a new folder if you do not have a folder yet.



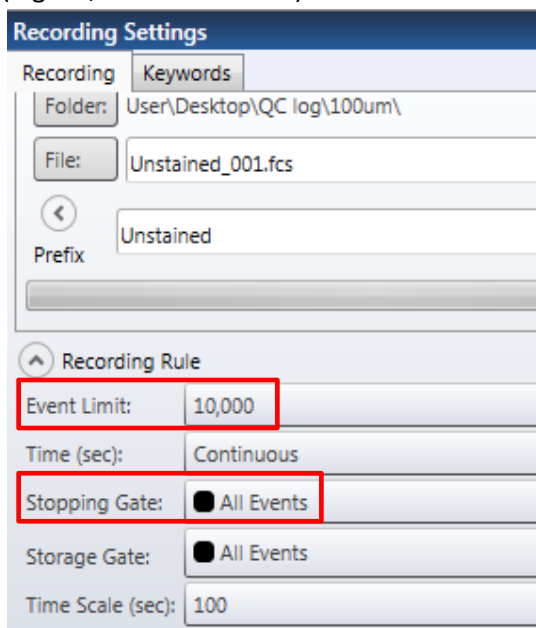
18. Key in the sample name in the **File** box.



The 'Recording Settings' dialog box is shown. The 'File' field is highlighted with a red box and contains 'Unstained_001.fcs'. The 'Folder' field contains 'User\Desktop\QC log\100um\'. The 'Prefix' field contains 'Unstained'.

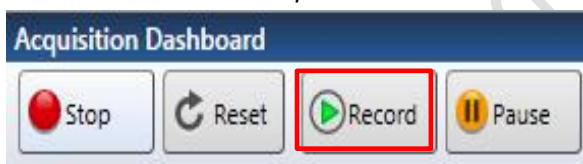
Imaging and Flow Cytometry Core

19. Set the number of events to record (**Event Limit**) and select the appropriate **stopping gate** (e.g. 10,000 events of P3).



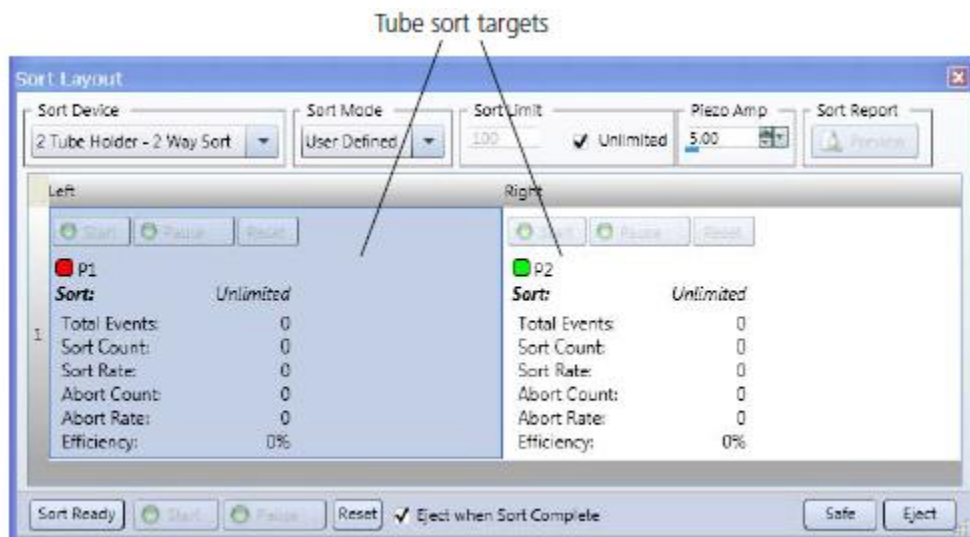
The image shows the 'Recording Settings' dialog box. The 'Recording' tab is selected. The 'Folder' is 'User\Desktop\QC log\100um\' and the 'File' is 'Unstained_001.fcs'. The 'Prefix' is 'Unstained'. The 'Recording Rule' section is expanded, showing 'Event Limit' set to '10,000', 'Time (sec)' set to 'Continuous', 'Stopping Gate' set to 'All Events' (indicated by a black circle), 'Storage Gate' set to 'All Events' (indicated by a black circle), and 'Time Scale (sec)' set to '100'. Red boxes highlight the 'Event Limit' and 'Stopping Gate' fields.

20. Click '**Record**' on the *Acquisition Dashboard*.



21. Right click at the corresponding tube position box (**Left/ Right** in the Diagram) in the *Sort Layout* pane. **Assign** the population to be collected.

Imaging and Flow Cytometry Core



22. Select the appropriate **Sort Mode**. Set a **Sort Limit** if needed by **unchecked Unlimited** and Key in the target value.



23. Press '**Plate**' to charge the deflection plates.

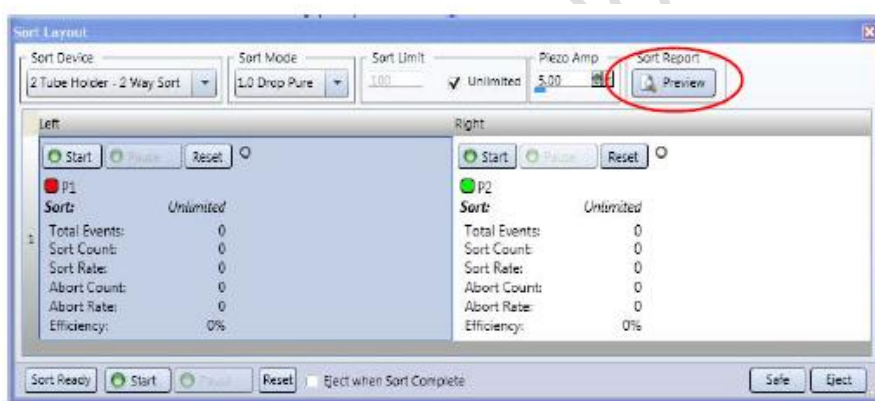
24. Click '**Sort Ready**' and '**Start**' in the Sort Layout pane to initiate cell sorting.

Imaging and Flow Cytometry Core



25. Click '**Pause**' if you want to pause the sort. The Sort Count will continue upon clicking '**Resume**'

26. Click '**Stop**' to complete your sort. Click '**Preview**' to view the **Sort Report**. Report can be saved as PDF file if needed.



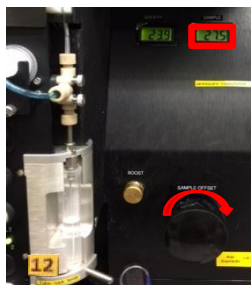
27. Click '**Stop**' on the *Acquisition Dashboard*. Unload your sample and press '**Backflush**'. Allow the flush for at least 5 seconds.

28. Continue with other samples or proceed to cleaning procedure.

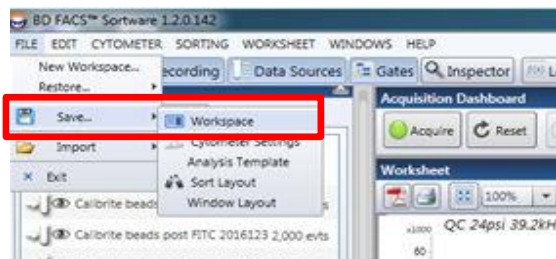
Imaging and Flow Cytometry Core

System Cleaning and Data Saving:

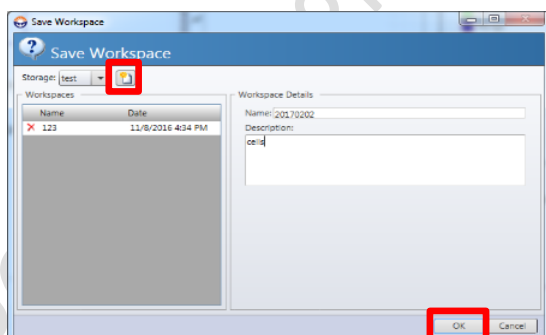
1. Load a tube of 3 ml FACS Clean Solution and set the sample pressure to 27.5 psi by adjusting the sample offset. Press **'Sample'** and let the solution run for **5 minutes**. Unload the tube and Press **'Backflush'**.
2. Repeat Step 1 with 3 ml FACS Rinse Solution and MilliQ water respectively. Leave the tube of water on the sample port when finish. DO NOT backflush after cleaning with water.



3. Click *File > Save > Workspace* to save workspace for next time.



Click the icon next to the drag list to **create a New Folder** for yourself. Fill in Workspace Details and click **'OK'** to save.



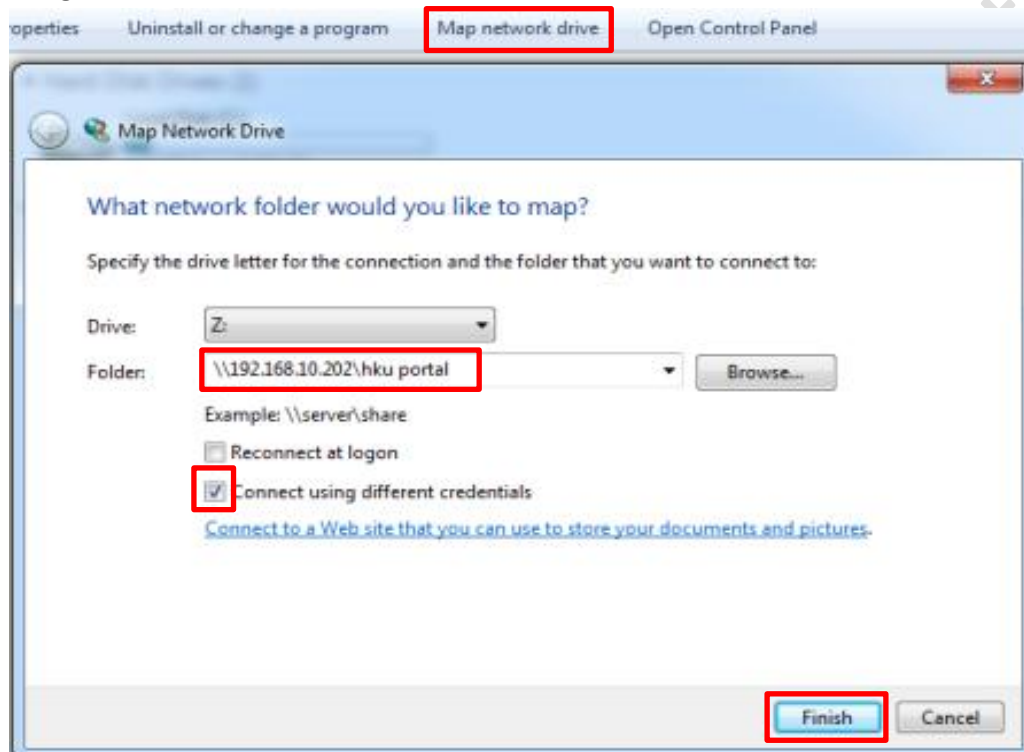
4. You may save PDF file by clicking **'PDF symbol'** for each sample and save it in your own folder in the **'User'** folder on **Desktop**.

Imaging and Flow Cytometry Core



Data Transfer:

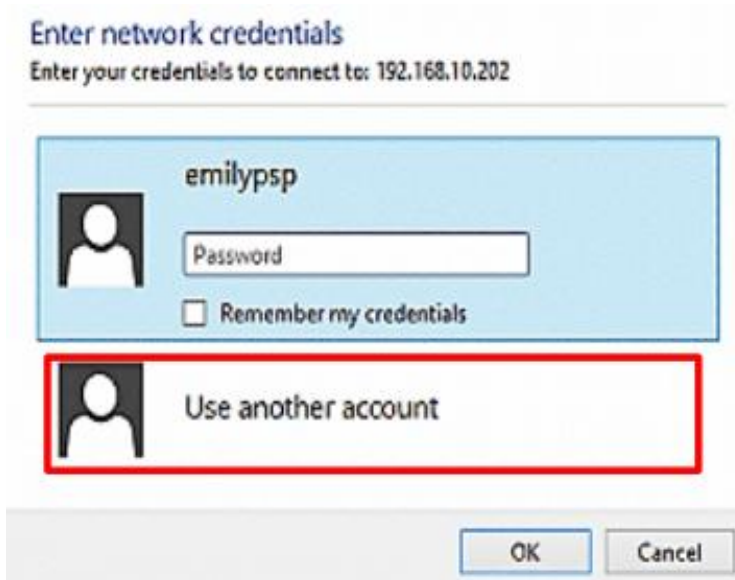
1. Double click 'Computer' icon on desktop.
2. Click 'Map Network Drive'. Key in \\192.168.10.202\HKU Portal ID at the Folder. Check 'Connecting using different credentials' and Click 'Finish'.



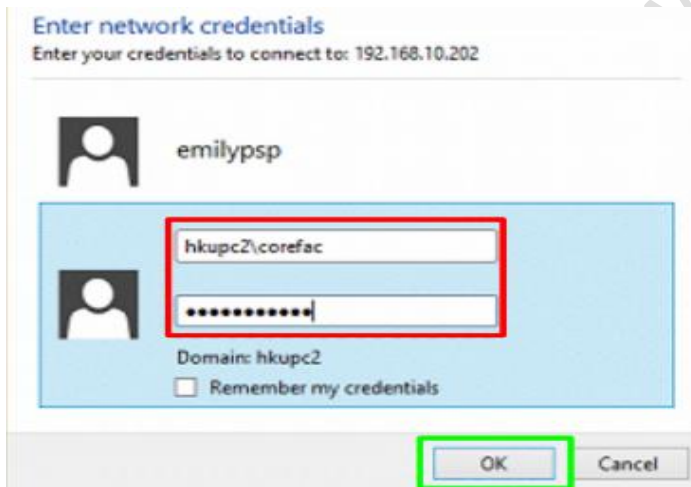
3. Another window would popup automatically. Click 'Use another account'.



Imaging and Flow Cytometry Core



4. Enter **hkupc2\HKU Portal ID** and your portal password. Click '**OK**'.



5. Copy and Paste your data from your folder to the server folder.
6. Disconnect the Network Drive when finish.



Imaging and Flow Cytometry Core

